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The predominant bacteria isolated from radicular cysts

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Abstract

Purpose: To detect predominant bacteria associated with radicular cysts and discuss in light of the literature.

Material and methods: Clinical materials were obtained from 35 radicular cysts by aspiration. Cultures were made from clinical materials by modern laboratory techniques, they underwent microbiologic analysis.

Results: The following are microorganisms isolated from cultures: *Streptococcus milleri Group* (SMG) (23.8%) [Streptococcus constellatus (19.1%) and Streptococcus anginosus (4.7%)], *Streptococcus sanguis* (14.3%), *Streptococcus mitis* (4.7%), *Streptococcus cremoris* (4.7%), *Peptostreptococcus pevotii* (4.7%), *Prevotella buccae* (4.7%), *Prevotella intermedia* (4.7%), *Actinomyces meyeri* (4.7%), *Actinomyces viscosus* (4.7%), *Propionibacterium propionicum* (4.7%), *Bacteroides capillosus* (4.7%), *Staphylococcus hominis* (4.7%), *Rothia denticariosa* (4.7%), *Gemella haemolysans* (4.7%), and *Fusobacterium nucleatum* (4.7%).

Conclusions: Results of this study demonstrated that radicular cysts show a great variety of anaerobic and facultative anaerobic bacterial flora. It was observed that all isolated microorganisms were the types commonly found in oral flora. Although no specific microorganism was found, Streptococcus spp. bacteria (47.5%) – especially SMG (23.8%) – were predominantly found in the microorganisms isolated. Furthermore, radicular cysts might be polymicrobial originated. Although radicular cyst is an inflammatory cyst, some radicular cyst fluids might be sterile.

Keywords: Radicular cyst, Bacteria, Predominant, Culture

Introduction

Radicular cysts are chronic inflammatory lesions [1] and are considered to develop as a result of inflammation in periapical of tooth with pulp necrosis [2]. They are the most common seen odontogenic cysts (approximately 50% of all odontogenic cysts) [3,4]. Many studies have been performed to determine the relationship between the periapical lesions and the microorganisms [5-7].

Iatrou et al. [8] performed a study on infected jaw cysts and obtained cyst fluid samples and cultures, reporting that there were different bacteria in cyst fluids. However, a sufficient number of studies have not performed to illuminate microbiological content of radicular cysts.

Infections are formed in periapical region as a result of odontogenic diseases, and unsuccessful dental treatments, can sometimes become serious intracranial infections [9,10] and common head–neck infections [11-13]. Sometimes,

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¹Department of Oral and Maxillofacial Surgery, Faculty of Dentistry, Abant Izzet Baysal University, Bolu, Turkey serious and intractable infections can be formed as a result of an acute exacerbation in radicular cysts [14,15]. Obtaining further information on microorganisms causing infection will make the disease easier to treat. Although treatment of the radicular cyst is generally performed as a surgical procedure, use of antibiotics is sometimes considered to be supplementary, or to eliminate or limit the bacterial pathogens. Therefore, better knowledge of microorganisms will help improve the selection of antibiotics in oral and maxillofacial surgery practices.

The purpose of this study was to investigate the microbiological cultures created with sample materials obtained from radicular cysts through modern laboratory techniques, detect predominant bacteria associated with these lesions, and discuss findings in light of the literature.

Materials and methods

We included patients who presented at Ondokuz Mayıs University, Faculty of Dentistry, Department of Oral and Maxillofacial Surgery Clinics. After examination and necessary tests, a total of 35 patients were randomly selected



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(25 males and 10 females) who were diagnosed with radicular cyst or residual cyst in maxilla and mandible, and clinically and radiologically delivered sample materials from cyst fluid to form microbiological cultures. The patients with antibiotics medication history within the last month were excluded from the study. All patients included in the study were asked to sign an informed consent form in accordance with Helsinki Declaration. The materials obtained from the patients were referred to SUVAM-TPL Bacteriology Department of Ondokuz Mayıs University for microbiological analysis.

Obtaining sample material from cysts and transporting to laboratory

To obtain cyst fluid from 35 patients who were diagnosed with radicular cyst (23) or residual cysts (12), buccal mucosa was wiped with a sponge using an antiseptic solution Isosol[®] (OTC İstanbul İlaç Paz. Med. İtr. İth. İhr. Ltd. Şti., Istanbul, TURKEY) containing povidone-iodine of 10% and then irrigated with 20 ml sterile normal saline using a 10 ml disposable sterile injector. The buccal mucosa space was cleaned with a sterilized aspirator tip concurrently. Asepsis was ensured to prevent bacterial contamination during collection of clinical material. The area into which the cyst expanded further, where fluctuation was detected, or where the bone was resorbed the most, was wiped to dry with a sterilized dry sponge. Approximately 1-1.5 ml of cyst fluid was obtained from the said area with a 10 ml disposable sterile injector inserted into the cyst cavity. The air in the injector was deflated completely, and the tip of the injector was bent to prevent air intake. The clinical materials obtained were transported to SUVAM-TPL Bacteriology Department of Ondokuz Mayıs University within 20 minutes. For a final diagnosis of cysts, the cyst mucosa obtained after the enucleation or marsupialization of cysts was transported to the pathology laboratory in a bottle containing 10% formalin [8].

Microbiologic laboratory procedures

After the sample materials in the injector were transported to the bacteriology laboratory, they were macroscopically checked by color, consistency, smell, and whether they contained sulfide granules. Blood agar was used to obtain aerobic cultures of clinical samples; Schaedler Agar with addition of vitamin K1, prepared daily, and broth (Oxoid LTD, Basingstoke, Hampshire, England) were used to obtain anaerobic cultures. Two different methods were used to have an anaerobic environment: The first one was closable 2.5-liter glass jars in which GENbag anaer (bioMérieux sa, France) used for 2.5 liter was placed to ensure an anaerobic environment. The second method was closable nylon bags in which bagcompatible GENbag anaer (bioMérieux sa, France) was placed to ensure an anaerobic environment. Strips

impregnated with methylene blue (Merk KGaA, Germany) were used to checker whether anaerobic conditions were formed in the atmosphere of both systems.

Aerobic cultures were incubated at 37°C for 24–48 hours, and anaerobic cultures were incubated at 37°C for 7 days. The colonies grown in Schaedler agar, which were incubated anaerobic, were compared with the colonies in blood agar, which was kept in aerobic conditions.

If there were haze or malodor in the Schaedler broth, just the colonies grown in anaerobic conditions, as a result of passaging, suggested anaerobic bacteria, whereas bacteria grown in either of them suggested facultative anaerobic bacteria. The isolated bacteria were stained with Gram's method. API 20 A panel (bioMérieux sa, France) and a computer system were used to identify the isolated bacteria [7]. The data obtained were evaluated statistically.

Review Board of Institute of Health Sciences in Ondokuz Mayis University approved this study. Reference number: (2004/ADÇ.119).

Results

The average age of all patients included in the study was 36.4 ± 14.1 years; the average age of males was 38.2 ± 15.3 years, and the average age of females was 32.0 ± 10.0 years. The clinical materials of 35 patients with cyst (23 radicular cysts and 12 residual cysts) that were obtained and cultured were analyzed microbiologically by aerobic and anaerobic. The bacteria were isolated in 13 (37.1%) of all cases. Sixteen different types of microorganisms were derived from 13 cases. From each case with bacteria grown, 1–3 bacteria (mean 1.61) were derived.

The following were microorganisms isolated from cultures: SMG (23.8%) [Streptococcus constellatus (19.1%) and Streptococcus anginosus (4.7%)], Streptococcus sanguis (14.3%), Streptococcus mitis (4.7%), Streptococcus cremoris (4.7%), Peptostreptococcus pevotii (4.7%), Prevotella buccae (4.7%), Prevotella intermedia (4.7%), Actinomyces meyeri (4.7%), Actinomyces viscosus (4.7%), Propionibacterium propionicum (4.7%), Bacteroides capillosus (4.7%), Staphylococcus hominis (4.7%), Rothia denticariosa (4.7%), Gemella haemolysans (4.7%), and Fusobacterium nucleatum (4.7%).

Of microorganisms isolated from the cultures, 14 were facultative anaerobic and 7 were obligate anaerobic bacteria. All facultative anaerobic bacteria were Gram positive; of obligate anaerobic bacteria, 3 types were Gram positive and the other 4 types were Gram negative (Table 1).

The status of isolated bacteria from the cultures are seen according to the localization of the cyst (Table 2), the properties of the cyst fluid (Table 3), and oral hygiene of the patients (Table 4).

The cyst mucosa obtained after surgical treatment of all patients diagnosed with radicular cyst or residual cyst,

Species of bacteria	Number of isolates	Gram stain	
Facultative anaerobes, microaerophiles, and capnophiles			
Streptococcus constellatus (SMG)*	4	Gram (+)	
Streptococcus anginosus (SMG)*	1	Gram (+)	
Streptococcus sanguis	3	Gram (+)	
Streptococcus mitis	1	Gram (+)	
Streptococcus cremoris	1	Gram (+)	
Staphylococcus hominis	1	Gram (+)	
Rothia denticariosa	1	Gram (+)	
Actinomyces viscosus	1	Gram (+)	
Gemella haemolysans	1	Gram (+)	
Total Aerobic Bacteria	14		
Strict anaerobes			
Prevotella buccae	1	Gram (-)	
Prevotella intermedia	1	Gram (-)	
Propionibacterium propionicum	1	Gram (+)	
Bacteroides capillosus	1	Gram (-)	
Actinomyces meyeri	1	Gram (+)	
Peptostreptococcus pevotii	1	Gram (+)	
Fusobacterium nucleatum	1	Gram (-)	
Total Strict Anaerobic Bacteria	7		

*SMG: Streptococcus milleri Group.

clinically and radiologically, was examined histopathologically and diagnoses were verified.

Discussion

Radicular cysts arise from periapical granulomas formed on devital teeth. They are the most common lesions [16,17] among tooth-originated periapical lesions, apart from periapical abscess [16] and periapical granulomas [17]. In addition, radicular cysts are the most common among all jaw cysts [3,4]. Therefore, determination of microorganisms associated with radicular cyst will ensure more conscious treatment of infections that might occur in the jaw.

The biggest problem with microbiological studies relating to the lesions in the periapical region is

Table 2 The relationship between the bacterial growing and the localization of radicular cysts

Localization of cyst	Bacterial growing		Case	
	Positive	Negative	number	
Maxilla	6	12	18	
Mandible	7	10	17	
Case Number	13	22	35*	

*Total number of the samples used in this study.

contamination with the microbial flora of surrounding tissue [5,7,8]. To avoid bacterial contamination in our study, the oral mucosa and teeth were wiped with povidone–iodine 10% prior to the aspiration of cyst fluid. The aspiration area was then irrigated with sterilized normal saline and wiped with a sterile sponge.

Antiseptic mouthwashes [18] and antibiotics is used to reduce the number of microorganisms [19]. In the case of administration of antibiotics, it is reported that antibiotics penetrated into the periapical lesions [20] and radicular cysts [21] in jaws. Iatrou et al. [8] isolated bacteria in 25 patients (67.6%) of 37 patients who had been on antibiotic medication before collecting samples from infected jaw cysts (1–6 days prior). The bacteria were grown in 12 patients (70%) of 17 patients who had not taken any antibiotics within the prior month. No statistical significant difference was found between the two groups; however, the patients who had taken any antibiotics within the prior month or had been on antibacterial medication were not included in our study because of the possibility of affecting the results.

Additionally, the results of this study showed that bacterial growing was higher in the cysts of mandible (53.85%) when compared with the cysts of maxilla (46.15%). We found that the transparent colored fluids (totally 9 cases) and the brownish colored viscous fluids (totally 3 cases) were been sterile and the bacteria was isolated from 57.89% of all straw colored fluids (totally 19 cases), 50% of the whitish colored fluids (totally 2 cases), and 50% the straw colored viscous fluids (totally 2 cases). Furthermore, we also found that positive culture was been in 33.33% of the patients who have well oral hygiene (totally 3 patients), in 30% of mild oral hygiene (totally 20 patients), and in 50% of poor oral hygiene (totally 12 patients). According to these results, oral hygiene, localization of cyst, and color of cyst fluid might be affect on bacterial growing.

In a study performed by Iatrou et al. [8] on infected jaw cysts – radicular, residual, dentigerous, globulomaxillary, incisive canal, and keratocyst – bacteria were isolated in 78.6% of microbiological cultures in 54 infected cyst fluid

Table 3 The relationship between the bacterial grow	ving
and the properties of cyst fluids	

Properties of cyst fluid	Bacterial growing		Case
	Positive	Negative	number
Transparent colored fluid	-	9	9
Straw colored fluid	11	8	19
Whitish colored fluid	1	1	2
Straw colored viscous fluid	1	1	2
Brownish colored viscous fluid	-	3	3
Case Number	13	22	35*

*Total number of the samples used in this study.

Table 4 The relationship between the bacterial growingand the oral hygiene of patients

Oral hygiene	Bacteria growing		Case
	Positive	Negative	number
Well	1	2	3
Mild	6	14	20
Poor	6	6	12
Case Number	13	22	35*

*Total number of the samples used in this study.

samples. The patients in our study were selected randomly and not categorized by infection. The bacteria were isolated in 13 (37.1%) of 35 cysts (23 radicular, 12 residual). This difference in results might be the selection of patients.

It is reported in the literature that the type of anaerobic bacteria were higher than the type of aerobe and facultative anaerobic bacteria when reviewing microbiological analysis of dentoalveolar infections [22-26]. Iatrou et al. [8] determined in their study performed on infected jaw cysts that a large part of isolated bacteria (89.2%) was anaerobic bacteria. Aerobe and facultative anaerobic bacteria growth were seen in 10.8% of the cases. In their study, a large part of isolated bacteria were found to be of those in normal oral microflora. For all that, in this study, of the microorganisms isolated from sample cultures, 14 (66.7%) were facultative anaerobic and 7 (33.3%) were obligate anaerobic bacteria. The isolated bacteria were determined to be the elements of normal oral flora.

In their study, latrou et al. [8] obtained a total of 84 types of bacteria in infected jaw cysts in which bacterial growth was observed. From each case with bacteria positive, 1–4 different colonies of bacteria were isolated. The greater part of isolated bacteria was Gram positive (59.5%) and the others were Gram negative (40.5%). In our study, 16 different types of bacteria were derived from 13 cases. Of the bacteria positive cases, 6 were polymicrobial and 7 were single types of bacteria. From each case with bacteria positive, 1–3 (mean 1.6) different types of bacteria were isolated. A large part of all isolated bacteria was Gram positive (81%), and the others were Gram negative (19%). There is a lot of diversity in the organisms found, the reason of these results may be caused by oral cavity organisms.

SMG has also been reported to be significant pathogens [27]. However, these pathogens are part of the normal flora of human oral cavity [28] and other mucous membranes [29-32]. SMG is not included in the approved lists of bacterial names, and is composed of three distinct species: *Streptococcus anginosus, Streptococcus constellatus,* and *Streptococcus intermedius* [33]. Further, SMG describes a group of anaerobic or microaerophilic streptococci. These organisms are frequently isolated from purulent infections of the head and neck region [34] and internal organs including: brain [35], liver [36], and lungs

[37]; and from cases of endocarditis [38] and meningitis [39]. They might cause infective endocarditis, such as streptococci of other mucous membranes; however, they have also been recurrently associated with severe suppurative infections, unlike streptococci of other mucous membranes [32].

SMG bacteria are important pathogenic microorganisms for infections and abscess of the orofacial region, according to literature survey. In the present study, SMG (23.8%) and *Streptococcus sanguis* (14.28%) were predominantly isolated from sample cultures. In addition, facultative anaerobic bacteria found to be higher in bacteria were isolated from cultures. Iatrou et al. [8] reported that the predominant group of bacteria isolated from infected jaw cysts was Gram-positive anaerobic coccus (36.9%). Hrvacanin et al. [40] analyzed radicular cyst fluid and often isolated *~-haemolytic streptococcus, Streptococcus pneumoniae, Staphylococcus epidermidis, and Streptococcus group B* type of bacteria.

Actinomyces are Gram-positive, non-acid fast, anaerobic or microaerophilic filamentous branched bacteria that are very difficult to grow in culture. In man, Actinomyces spp. bacteria may cause severe head and neck infections. The pathogenic Actinomyces most frequently isolated is Actinomyces israelii. Actinomyces propionica, Actinomyces naeslundii, Actinomyces viscosus, and Actinomyces odontolyticus are less common infections. These bacteria are all normal commensals of the human oral cavity [41-46]. Actinomyces spp. bacteria were not isolated by Iatrou et al. [8]. However, Hirsberg et al. [47] performed a study and analyzed periapical biopsy samples histologically. They found typical actinomycotic colonies in 1.8% of the samples. Additionally, Tseng at al. [15] reported the presence of Actinomyces spp. bacteria in the case of a radicular cyst. The type of Actinomyces meyeri and Actinomyces viscosus (9.52%) bacteria were isolated also in our study.

Additionally, radicular cyst is generally treated using surgical enucleation or/and marsupialization. If it becomes necessary to antibiotic therapy to help for the treatment of radicular cyst, the surgeon should consider the results of this study for the selection of antibiotic.

Conclusion

Results of this study demonstrated that radicular cysts show a great variety of anaerobic and facultative anaerobic bacterial flora. It was observed that all isolated microorganisms were the types commonly found in oral flora. Although no specific microorganism was found, Streptococcus spp. bacteria (47.5%) – especially SMG (23.8%) – were predominantly found in the microorganisms isolated. Furthermore, radicular cysts might be polymicrobial originated. Although radicular cyst is an inflammatory cyst, some radicular cyst fluids might be sterile.

Consent statement

All patients included in the study were asked to sign an informed consent form in accordance with Helsinki Declaration.

Abbreviations

SMG: Streptococcus Milleri Group; Spp: Species.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

MT and MM conceived of the study, and participated in its design and coordination and helped to draft the manuscript. MT, IS, and CB collected the data. MTo performed microbiological examination, analysed and interpreted the data. MT, HOK and SE wrote the manuscript. HOK and SE participated in its English editing. All authors read and approved the final manuscript.

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References

- Marcal JR, Samuel RO, Fernandes D, de Araujo MS, Napimoga MH, Pereira SA, Clemente-Napimoga JT, Alves PM, Mattar R, Rodrigues V Jr, et al: T-helper cell type 17/regulatory T-cell immunoregulatory balance in human radicular cysts and periapical granulomas. J Endod 2010, 36(6):995–999.
- Nair PN, Sundqvist G, Sjogren U: Experimental evidence supports the abscess theory of development of radicular cysts. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2008, 106(2):294–303.
- Avelar RL, Antunes AA, Carvalho RW, Bezerra PG, Oliveira Neto PJ, Andrade ES: Odontogenic cysts: a clinicopathological study of 507 cases. J Oral Sci 2009, 51(4):581–586.
- Sharifian MJ, Khalili M: Odontogenic cysts: a retrospective study of 1227 cases in an Iranian population from 1987 to 2007. J Oral Sci 2011, 53(3):361–367.
- Abou-Rass M, Bogen G: Microorganisms in closed periapical lesions. Int Endod J 1998, 31(1):39–47.
- Samaranayake LP, Stassen LF, Still DM: A microbiological study of pre- and postoperative apicoectomy sites. Clin Oral Investig 1997, 1(2):77–80.
- Iwu C, MacFarlane TW, MacKenzie D, Stenhouse D: The microbiology of periapical granulomas. Oral Surg Oral Med Oral Pathol 1990, 69(4):502–505.
- Iatrou IA, Legakis N, Ioannidou E, Patrikiou A: Anaerobic bacteria in jaw cysts. Br J Oral Maxillofac Surg 1988, 26(1):62–69.
- Corson MA, Postlethwaite KP, Seymour RA: Are dental infections a cause of brain abscess? Case report and review of the literature. Oral Dis 2001, 7(1):61–65.
- Jimenez Y, Bagan JV, Murillo J, Poveda R: Odontogenic infections. Complications. Systemic manifestations. Medicina oral, patologia oral y cirugia bucal 2004, 9:143–147. 139–143.
- Brunworth J, Shibuya TY: Craniocervical necrotizing fasciitis resulting from dentoalveolar infection. Oral Maxillofac Surg Clin North Am 2011, 23(3):425–432.
- Rega AJ, Aziz SR, Ziccardi VB: Microbiology and antibiotic sensitivities of head and neck space infections of odontogenic origin. J Oral Maxillofac Surg 2006, 64(9):1377–1380.
- Baqain ZH, Newman L, Hyde N: How serious are oral infections? J Laryngol Otol 2004, 118(7):561–565.
- Hirvonen TP, Ertama L: [An infected radicular cyst--a rare cause for facial cellulitis]. Duodecim; laaketieteellinen aikakauskirja 1998, 114(17):1734–1736.

- 15. Tseng SK, Tsai YL, Li UM, Jeng JH: Radicular cyst with actinomycotic infection in an upper anterior tooth. *J Formos Med Assoc* 2009, **108**(10):808–813.
- Ramachandran Nair PN, Pajarola G, Schroeder HE: Types and incidence of human periapical lesions obtained with extracted teeth. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 1996, 81(1):93–102.
- 17. Becconsall-Ryan K, Tong D, Love RM: Radiolucent inflammatory jaw lesions: a twenty-year analysis. Int Endod J 2010, 43(10):859–865.
- Metin M, Tek M, Sener I: Comparison of two chlorhexidine rinse protocols on the incidence of alveolar osteitis following the surgical removal of impacted third molars. J Contemp Dent Pract 2006, 7(2):79–86.
- 19. Mombelli A: Antimicrobial advances in treating periodontal diseases. *Front Oral Biol* 2012, **15:**133–148.
- Akimoto Y, Kaneko K, Fujii A, Yamamoto H: Ampicillin concentrations in human radicular granuloma following a single oral dose of bacampicillin. J Oral Maxillofac Surg 1992, 50(1):11–13.
- Akimoto Y, Mochizuki Y, Uda A, Shibutani J, Nishimura H, Komiya M, Kaneko K, Fujii A, Yamamoto H: Ampicillin concentrations in radicular cysts following a single oral administration of bacampicillin. *Gen Pharmacol* 1993, 24(4):895–898.
- Lewis MA, MacFarlane TW, McGowan DA: Quantitative bacteriology of acute dento-alveolar abscesses. J Med Microbiol 1986, 21(2):101–104.
- von Konow L, Nord CE, Nordenram A: Anaerobic bacteria in dentoalveolar infections. Int J Oral Surg 1981, 10(5):313–322.
- 24. Labriola JD, Mascaro J, Alpert B: The microbiologic flora of orofacial abscesses. J Oral Maxillofac Surg 1983, 41(11):711–714.
- 25. Sklavounos A, Legakis NJ, Ioannidou H, Patrikiou A: Anaerobic bacteria in dentoalveolar abscesses. *Int J Oral Maxillofac Surg* 1986, **15**(3):288–291.
- 26. Williams BL, McCann GF, Schoenknecht FD: Bacteriology of dental abscesses of endodontic origin. J Clin Microbiol 1983, 18(4):770–774.
- 27. Whiley RA, Beighton D, Winstanley TG, Fraser HY, Hardie JM: **Streptococcus intermedius**, **Streptococcus constellatus**, **and Streptococcus anginosus** (the Streptococcus milleri group): association with different body sites and clinical infections. *J Clin Microbiol* 1992, **30**(1):243–244.
- Mejare B, Edwardsson S: Streptococcus milleri (Guthof); an indigenous organism of the human oral cavity. Arch Oral Biol 1975, 20(11):757–762.
- Poole PM, Wilson G: Occurrence and cultural features of Streptococcus milleri in various body sites. J Clin Pathol 1979, 32(8):764–768.
- Ruoff KL, Kunz LJ: Identification of viridans streptococci isolated from clinical specimens. J Clin Microbiol 1982, 15(5):920–925.
- 31. Unsworth PF: The isolation of streptococci from human faeces. *J Hyg (Lond)* 1980, **85**(1):153–164.
- Gossling J: Occurrence and pathogenicity of the Streptococcus milleri group. Rev Infect Dis 1988, 10(2):257–285.
- Approved Lists of Bacterial Names (Amended). Edited by Skerman VBD, McGowan V, Sneath PHA. Washington (DC): ASM Press; 1989.
- Han JK, Kerschner JE: Streptococcus milleri: an organism for head and neck infections and abscess. Arch Otolaryngol Head Neck Surg 2001, 127(6):650–654.
- Kowlessar PI, O'Connell NH, Mitchell RD, Elliott S, Elliott TS: Management of patients with Streptococcus milleri brain abscesses. J Infect 2006, 52(6):443–450.
- Chua D, Reinhart HH, Sobel JD: Liver abscess caused by Streptococcus milleri. Rev Infect Dis 1989, 11(2):197–202.
- Wong CA, Donald F, Macfarlane JT: Streptococcus milleri pulmonary disease: a review and clinical description of 25 patients. *Thorax* 1995, 50(10):1093–1096.
- Concistre G, Chiaramonti F, Miceli A, Glauber M: Mitral and aortic valve endocarditis caused by a rare pathogen: Streptococcus constellatus. Interact Cardiovasc Thorac Surg 2012, 14(6):889–890.
- Tecson-Tumang F, Sen P, Kapila R: Fatal Streptococcus MG-intermedius (Streptococcus milleri) meningitis in an adult. Am J Clin Pathol 1982, 77(4):480–484.
- 40. Hrvacanin S: [Bacteriologic study of the contents of radicular cysts]. *Medicinski pregled* 2002, **55**(1–2):41–43.
- 41. Smego RA Jr, Foglia G: Actinomycosis. Clin Infect Dis 1998, 26(6):1255–1261. guiz 1262–1253.
- 42. Belmont MJ, Behar PM, Wax MK: Atypical presentations of actinomycosis. Head Neck 1999, 21(3):264–268.
- Aguirrebengoa K, Romana M, Lopez L, Martin J, Montejo M, Gonzalez De Zarate P: [Oral and cervicofacial actinomycosis. Presentation of five cases]. Enfermedades infecciosas y microbiologia clinica 2002, 20(2):53–56.

- Bhargava D, Bhusnurmath B, Sundaram KR, Raman R, Al Okbi HM, Al Abri R, Date A: Tonsillar actinomycosis: a clinicopathological study. *Acta tropica* 2001, 80(2):163–168.
- 45. Stewart MG, Sulek M: Pediatric actinomycosis of the head and neck. *Ear Nose Throat J* 1993, **72**(9):614–616. 618–619.
- Volante M, Contucci AM, Fantoni M, Ricci R, Galli J: Cervicofacial actinomycosis: still a difficult differential diagnosis. Acta Otorhinolaryngol Ital 2005, 25(2):116–119.
- 47. Hirshberg A, Tsesis I, Metzger Z, Kaplan I: **Periapical actinomycosis: a clinicopathologic study.** *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2003, **95**(5):614–620.

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